

Characterization of a novel member of murine semaphorin family[☆]

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Abstract

Semaphorin gene family contains a large number of secreted and transmembrane proteins, and some of them are functioning as the repulsive and attractive cues of the axon guidance during development. Here we report murine orthologues of a novel member of class 6 semaphorin gene, semaphorin 6D (Sema6D), mapped on the chromosome 2. Sema6D is mainly expressed in the brain and lung, and the ubiquitous expression in the brain continues from embryonic late stage to adulthood, as determined by Northern blot and in situ hybridization. We also found that Sema6D has five different splicing variants, and the expression patterns of individual isoforms differ depending on the tissues. Thus, Sema6D may play important roles in various functions including the axon guidance during development and neuronal plasticity.

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During embryogenesis, axons reach their specific targets correctly to form the complex neural network found in the mature functional nervous system. Several groups of axon guidance molecules such as semaphorins, ephrins, netrins, and slits have been reported to repel or attract the growing axons that express their cognate receptors [1].

Semaphorins are secreted and transmembrane proteins containing a conserved domain (semaphorin domain) of about 500 amino acids are found in both vertebrates and invertebrates [2,3]. So far, more than 20 kinds of semaphorin genes have been identified and classified into seven classes and a virus semaphorin [4]. Among them, semaphorin 3A (Sema3A) is the first identified semaphorin in vertebrates on the basis of its ability to induce the collapse of axonal growth cones of dorsal root ganglion (DRG) [5]. Sema3A-deficient mice

showed a severe abnormality in the axonal projection pattern in the peripheral nervous system during embryogenesis [6]. Neuropilins are functional receptors for class 3 semaphorins and plexins are also known as receptors for other types of semaphorins [7–15]. Moreover, semaphorin 4D/CD100 functions in the immune system and semaphorin 3C (Sema3C) functions during cardiac development [16,17]. Thus, semaphorin family genes perform many important biological functions besides the axon guidance.

To identify a novel semaphorin gene, we searched human expressed sequence tag (EST) databases and identified a novel human semaphorin cDNA, KIAA1479. By using KIAA1479 as a probe, here, we cloned a novel murine transmembrane semaphorin gene, semaphorin 6D (Sema6D), and its four splicing variants. Northern blot and in situ hybridization analyses suggest their important roles in the brain development and functions.

Materials and methods

Identification and cloning of semaphorin 6D. We searched human EST databases and identified a novel human semaphorin cDNA,

[☆] Abbreviations: aa, amino acid; bp, base pair; DRG, dorsal root ganglion; E, embryonic day; EST, expressed sequence tag; GAPDH, glyceraldehyde-3-phosphate dehydrogenase; ORF, open reading frame; RT, reverse transcriptase; Sema6D, semaphorin 6D; SSC, standard saline citrate; VEGF, vascular endothelial growth factor.

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KIAA1479, which is a member of class 6 semaphorin and thus was named semaphorin 6D (SEMA6D) according to the rule of Semaphorin Nomenclature Committee [4]. To clone the murine semaphorin 6D (Sema6D) cDNA, adult mouse (C57BL/6) brain cDNA library was screened with a 1.5 kb *HincII* fragment of human SEMA6D cDNA as a probe. Hybridization was performed in $6\times$ SSC, $5\times$ Denhardt's solution, 0.2% SDS, and 100 μ g/ml herring sperm DNA at 55°C overnight. The filters were washed with $2\times$ SSC–0.2% SDS four times at 55°C. The positive clones were sequenced by use of an ABI PRISM 3100 Genetic Analyzer (Applied Biosystems, CA, USA). Thus, a 6330 bp Sema6D-1 cDNA containing the open reading frame and four isoforms were obtained. The accession numbers of Sema6D-1, Sema6D-2, Sema6D-5, Sema6D-6, and Sema6D-4 are deposited as AB091532, AB091533, AB091534, AB091535, and AB091536, respectively.

RT-PCR. Reverse transcriptase (RT) reaction was performed by ThermoScript RT (Invitrogen, CA, USA) with 2 μ g of total RNA. PCR was performed by TaKaRa Ex Taq (TaKaRa, Japan) with an exon 15 primer TTAAGCCAGGGAGTTTGTGAGAGA and an exon 19 primer TTCATGTGGACCATCTGATTGGAT. PCR products were electrophoresed on 4% agarose gel.

Northern blot hybridization. Northern blot membranes purchased from Seegene, (Seoul, Korea) were used. Hybridization was performed in 50% formamide, $6\times$ SSC, $5\times$ Denhardt's solution, 0.2% SDS, and 100 μ g/ml herring sperm DNA at 42°C overnight. The filters were washed with $2\times$ SSC–0.2% SDS three times and $0.2\times$ SSC–0.2% SDS once at 65°C. A 2.5 kb *BanII*–*BglII* fragment of Sema6D-1 cDNA was used as a probe.

In situ hybridization. In situ hybridization was performed as described [18]. Anesthetized ICR mice were perfusion-fixed with 4% paraformaldehyde and dissected organs were sectioned after paraffin embedding. After in situ hybridization staining, the sections were counterstained with Kernechtrot stain solution (Muto Pure Chemicals, Tokyo, Japan). To generate probes, Sema6D-1 fragment (2317–2886, 570 bp) was cloned. Digoxigenin-labeled riboprobes were prepared by transcription of linearized plasmids using T7 or T3 RNA polymerase and DIG RNA labeling kit (Roche Molecular Biochemicals, Mannheim, Germany). Sense probes were used as controls.

Results

Identification and cloning of human and murine semaphorin 6D genes

We identified one novel transmembrane semaphorin cDNA, KIAA1479. Sequence analysis revealed that KIAA1479 contained a predictable open reading frame (ORF) and was a novel member of class 6 semaphorin (Fig. 1B). KIAA1479 was named semaphorin 6D (SEMA6D) according to the rule of Semaphorin Nomenclature Committee [4]. SEMA6D encodes 1011 amino acid (aa) protein with a predicted molecular weight of about 113 kDa. Then, murine semaphorin 6D (Sema6D) was cloned by use of human SEMA6D cDNA as a probe (Fig. 1A). The length of Sema6D cDNA was 6330 bp and the ORF of Sema6D are 3033 bp (1011 aa). Murine Sema6D protein showed a 95.3% identity with human SEMA6D protein (Fig. 1B), and similarity to other class 6 semaphorins was: Sema6A1 (45% identity), Sema6B (44% identity), and Sema6C (38% identity) [19–22].

During the Sema6D cloning, we obtained four kinds of Sema6D isoforms. Because human SEMA6D was recently reported by others independently [23], these Sema6D and isoforms were renamed Sema6D-1 (1011 aa predictable protein), Sema6D-2 (998 aa), Sema6D-5 (1030 aa), Sema6D-6 (1054 aa), and Sema6D-4 (1073 aa) according to human SEMA6D nomenclature. These isoforms were generated by alternative splicing (Fig. 2 and see below).

Genomic structure and chromosomal localization

Sema6D gene was mapped on chromosome 2 by the use of Celera mouse genomic database. Comparison between Sema6D cDNA and mouse genomic DNA sequences revealed that Sema6D gene is composed of at least 19 exons. Sema6D gene spans more than 56 kb of the genomic DNA. The first exon including 5' non-coding region was about 42 kb distant from the second exon that includes the initiation codon ATG. Thus, the length of other 18 exons and 17 introns of Sema6D gene is about 14 kb. In Sema6D isoforms, exon 16a (39 bp) was deleted in the ORF of Sema6D-2 cDNA, while exon 17 (57 bp) was added to Sema6D-5. Sema6D-6 lacked exon 16a and attached exon 18 (168 bp). Sema6D-4 did not have exon 16a but had exons 17 and 18 (Fig. 2).

Expression pattern

To clarify the temporal and regional expression of Sema6D, Northern blot hybridization was performed (Fig. 3). Sema6D transcript was about 6.5 kb in size, and the length was comparable to the size of the cloned Sema6D cDNA. Its expression during embryogenesis was analyzed (Fig. 3A). Sema6D was detected on embryonic day 10.5 (E10.5) and its expression continued until birth. It was expressed predominantly in adult brain and lung, moderately in heart, small intestine, skeletal muscle, uterus, and placenta (Fig. 3B). In lung and placenta, additional Sema6D transcripts were detected, but their natures remain to be elucidated. As Sema6D was expressed predominantly in adult brain, we analyzed the developmental changes in Sema6D expression of the brain (Fig. 3C) and its regional distribution in adult brain (Fig. 3D). Sema6D expression in the brain was detected from E17.5 to 12 months at similarly high levels. Furthermore, Sema6D was expressed ubiquitously in the brain with relatively higher expressions in the thalamus, hypothalamus, midbrain, cerebellum, pons and medulla oblongata, and spinal cord. To further study Sema6D expression patterns, in situ hybridization was performed (Fig. 4). At E13.5 a high Sema6D expression was detected in the neopallial cortex (Fig. 4B, shown in blue). In adult brain

A

CTCCGCTGACGCTCTGGGTGCGTGATGGGGGTGGGACCCGAGCGGGAGCCCGAGCGC 60
 AGCTGTGTGCGGACCGGTGTGTTTGGGTGGGCGTAGCTGCTAGCTCCCGGGAGTTCATT 120
 GCTCTCCCAAGCCACCGGTCTCCAGCGGTGCGGAGCGGCGGAGGCTTCTGGCTGAC 180
 CAGTCCGACGCTGCGCGGTGGGCGGCTGGGGGCGGGGCGGAGCTGCGGCTTCCTCT 240
 AGGCGACCCCTCTCTTGAAGCGCACCCCTAAGGAGAAAGAACACCTCTGGCTGCGAG 300
 GAAAGTTTCTTCTTGGCAGGCGCGCTGCGGCGGCTCGCAGGCGCTCAGGTGAGCG 360
 CCGCTTCTCTGCTGGGCGCTTGGAGCTTCCAGCGCGCGCGAGCGAGCGGCGCGCG 420
 GGGCGAGCTTCCGCGGGGAGAACGAGGAGCAGCGTGGGAGTGCGGAGCAGCTGACTGTC 480
 CACGCGGGGACCGTGAGCACCACTCGCGGCTTCCAGCCACTTCTCCCGCACTTTTG 540
 CGTGTCTTGTCTCTGGCTGAAAAATGCTCCACGAAGGCCATAGACCGGGTCTCTCT 600
 CAGCGGCGAAGGGCTCGGCGGGGAGCGGTGTTACGCTCTCAGCTTCCCATCAATATATGA 660
 TGGAAACAAATAAGGAAAGTCAATTTGTATGAGCGCGCTCTTTCGACGCGAGGCGCT 720
 CTTGTAGCTGGAGCAGGCGAGGCAACCAACCAACAGTAGCTTAAGTGGGAC 780
 TTCTGAGGAGGGGAGCTCAGATTCTCTTGGCCAACTCGGGGTCTCTCTGCTTTGG 840

M G F L L L W

TTCTGCGTGTCTCTCTGCTTCCAGGTTACGCGGCTCAGCTTCCCAAGAGCAGT 900
 F C C V L F L L V S R L R A V S F P E D D 27
 GAGCGCTCAACCGGTGACTACTACTTCAAGCAATATCCGTTTATAGAGCGC 960
 E P L N T V D Y H Y S R Q Y V P F R G R 47
 CTTTCAGGCAACAGTCCGACGAGCTGAGCTTTCAGCTGTGTAATTTAGAGAC 1020
 P S G N E S Q H R L D F Q L M L K I R D 67
 ACATCTTATATCTCGCGAGGATAGCTATATAGCTGAATTTAAATGAAATCCCCAA 1080
 T L Y I A G R D Q V Y T V N L N E I P 87
 ACAGAGGTGATACCAAGCAAGCTGACGCTGGAGGTCCAGACAGCAGGATGAGAAAAAT 1140
 T E V I P S K K L T W R S R Q Q D R E N 107
 TGTGCTATGAAAGCAAGCTAAGATGAAAGTCCCAACTCATCAAGCTTTTGTGCCA 1200
 C A M K K K H K D E A C H N F I K V F V P 127
 AGAAATGATGAGATGGTTTGTCTGTGTGTAATGTTTCAACCCGATGTCGAGATAC 1260
 R N D E M V F V C G T N A F N P M C R Y 147
 TATAGTTGAGAAGCTTAGATGATGAGGGAAGAAATAGTGGCTGGCAGGATCCCG 1320
 Y R L R T L E Y D G G E I S G L A R C P 167
 TTTGATGCCCAACCAACCTGCGCCCTCTTTCGCTGATGAAACCTCTATTCTGCCACA 1380
 F D A R Q T N V A L F A D G K L Y S A T 187
 GTGGCTGATTTCTTGGCAGTGTGCTGATTTACAGAAGCATGGGAGATGATCTGCC 1440
 V A D F L A S D A V I Y R S M G D G S A 207
 CTTGCGCAACAAATACAGTTCGAAGTGGATCAAGAACACACACTCTCTCATGCGCAAT 1500
 L R T I K Y D S K W I K E P H F L H A I 227
 GAATATGGAACATGCTATCTTCTTCTCAGAGAAATCGCGGTGGAACATAAATACTTA 1560
 E Y G N Y V Y F F F R E I A V E H N L 247
 GGCAAGGCTGTGATTCGCGGTGGCTGCATTTGTAAAAACACATGGTGGCTCAGAG 1620
 G R A V I S R V A R I C K N D M G G S Q 267
 CGGCTCTGGAGAAACATGAGCTTCTTCTTAAAGCTCGGCTGACTGCTCGCTCTCT 1680
 R V L E K H W T S F L K A R L N C S V P 287
 GGAGATCTCTTTTCTACTGCGACTTCTGCGAGTCTATAACAGACATATCAAAATCAAT 1740
 G D S F F V G V F T Q S I T D I Q I M 307
 GGATATCCCACTGGTGGGTCTTCCACACAGCTCAACAGCATCTCTGTTCTGCA 1800
 G T P T V V V F T Q L N S I G S A 327
 GTCTGTGCTTTAGCATGAGACATGAGAAAGTGTCAAGGCGCTGCAAGAG 1860
 V C A F S M D D I E K V F K R F K E Q 347
 AAAACCCAGACTCTTTTGGACAGAGTCCGGAAGAACAGTACCAAAACCAAGGCTC 1920
 K T P D S V W T A V A P E D K V P K P R P 367
 GGCTGTGTGCAACAGGCTCGACAGCTTCAAGACCTTCATCGACTTTCAGAT 1980
 G C C A K K H G L A E A Y K T S I D F D 387
 GACACCTGGCTTTCATCAAGTCCACCGCTGATGAGCTTGCCTGCCACCTATGCC 2040
 D T L A F I K S H P L M D S A V P P I A 407
 GATGAGCCCTGGTTTCAAAAGACAGGCTGAGTACAGTTGCTGGCTGTAAGCTGGCGT 2100
 D E P W F T K T R V R Y R L T A I E V D 427
 CGTTTCAGAGGCGCATACCAACTACACAGTCTATCTTCTGCTGTAAGCTGGCGT 2160
 R S A G P Y Q N Y T V I F V G S E A G V 447
 GTACTTAAAGTTTGGCAAGACCATCTCTTCTCTGTAAGTACAGTGTATTAAGT 2220
 V L K V L L A K T S P F L N D S V L L E 467
 GAGATTAAGCTTATAACCGCAAGTACGAGCGGAGAGTGGAGGAGCAAGGTTG 2280
 E I E A Y N P A K C S A E S E E D R K V 487
 GTCTCATACAGCTGGACAGGATACCATGCTTTATACGTGGCTTCTCTAGTGGCGT 2340
 V S L Q L D K D H H A L Y V A F S C T V 507
 GTCCGCTACCCCTCAGCGCTGTGAGGCTGACGAGTGTGTAAGTCTTGTGATGCA 2400
 V R I P L T S R G E E R Y G S C K K S C I A 527
 TCAGTGACCCGCTACTGTGTGTTTAAAGCGGAGTGTGAGAGAGTGAACCTTAGG 2460
 S R D P Y C G W L S Q G V C E R V T L G 547
 ATGCTGCTGTAAACCGAAGCTTCTTGTCTTCCATAACACAGCCCTGGAGTATGAG 2520
 M L L L T E D F F A F H N H S P G G A E 567
 CAGGACAGGAGTACGGAACACAGCCACCTAGGGGAGTGCACGCTGTACGTTGGGAA 2580
 Q D T E Y G N T A H L G D C H G V R W E 587
 GTCCAGCTTGGAGAAATCCAGTGTGTCACATGAATGCTCTCATACCTGCGGTGTT 2640
 V Q S G E S N Q M V H M N V L I T C V F 607
 CGCGCTTTTGTCTGGGCGGCTTCTACGAGAGTGGCGCTGACTGCTACCGTGACATG 2700
 A A F V L G A F I A G V A V Y C Y R D M 627
 TTTGTTTCGGAAGACAGAAAGTCCATAAGACGACGAAATCCGCGCAGCTGTCACAG 2760
 F V R K N R K I H K D A E S A Q S C T D 647
 TCCAGCGGAAGCTTTCGCAAGACGCGCTTCTTGAAGGAAAGAACACCCCAATACAG 2820
 S S G S F A K L N L G L F D S P V K E Y Q 667
 CAGAACTTGTATCTCCCAACTCTACAGCAACCTGCTGACCACTCGGAAGGAACCTGCCA 2880
 Q N I D S P K L Y S N L L T S R K E L P 687
 CCAAAACAGGATACAAAGTCCATGGCGGTGAGACACAGAGGCGAGCTCCGAGCTGGCT 2940
 P N T D T K S M A V D H R G A Q P P E L A 707
 GCTCTCCCGAGCGGAATCCACACTGCTCTCCAGCAGAGCCCTGACGCGCATGAAG 3000
 A L P T P E S T P V L H Q K T L Q A M K 727
 AGCCACTCTGAGAAGGCCACAGCGCGCTTCTTGAAGGAAAGAACACCCCACTTTT 3060
 S H S E K A H S H G A S R K E H P Q F F 747
 CTTCTAGTCTCCACCCCTTCCCATGTAGTACGCGGATATCCCACTGCGCATGCT 3120
 P T S S P P P H S P L S H G H I P S A I V 767
 TTTCCAAAGCCGCTACGAGCTACATCACTCTTCCAACTGCAATGCCACAAAGGCC 3180
 L P N A T H D Y N T S F S N S N A H K A 787
 GAAAGAGAGCTTACAGATGATACCTCTTACAGAGTATCCAGTACGAGCGGAGGAC 3240
 E K K L Q G A T H D P L T K S S K R E H 807
 CGCGGCTGTGTGATTCAGGAATACCTCAATGATCTCTGGAAGCATCAATTAAGACCCA 3300
 R S V S D S R N T L N D L L K H L N D P 827
 AACAGTAACCCAAAGCCATCTGGGAGAGATCCATGGCTCATCAACACCTCATGCTG 3360
 N S N P K A I L G E I H M A H Q T L M L 847
 GACCCGTTGGGACCAATGGCTGAGGTCACCAAGGTCCTTACCGGGAGGCATCTTA 3420
 D P V G P M A E V P P K V P N R E A S L 867
 TACTTCCCTCCCTCCACATCCCGAAGATGCTCAACCAAGAGTAGATGCTCCCAAC 3480
 Y S P P S T L P R N S P T K R V D V P T 887

ACTCCTGGGGTGCCAACTGACTTCTCTGGAAGACAAAGGGGTATACAAAAATCTCTCC 3540
 T P G V P M T S L E R Q R G Y H K N S S 3907
 CAGAGGACTCTATATCTGCGTGCCTAAAGACTTAAACTCACCAGTGGTGTGTTTGTGA 3600
 Q R H S I S A V P K N L N S P N G V L L R 927
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 S R Q P S M N R G G Y M P T P T G A K V 947
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 D Y I Q G T P V S V H L Q P S L S R Q S 967
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 S Y T S N G T L P R T G L K R T P S L K 987
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 P D V P P K P S F V P Q T T S V R P L N 1007
 AAGTACAGTACTAGGCTCAAGTATGCTATTCCCGTGTGGCTTTATCTCTGCTCCGTG 3900
 K Y T Y * 1011
 TTGCGAGGAAGCTGTAGGGTACCTCAGATGAGATACCTGCTGTATTTTAAAGAAAC 3960
 AAGTAGCCAAAGAACTCTGTCACTTTGGTAACACAGAACTTGCACATGTAGCTACTA 4020
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 CGTTTGAAGCAGAGGAGTGTGAGCAGGAGGAGGCTCCCTTCAAGCTGTAAAGAGCTG 4140
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 AAGCAGTGTGTGAAATCAATCTTGACCCAACTGTAGCAATTTTATTTCTTGAGT 4380
 GTGGCACTGTGTGTTTATTTTGTGTTTGTGTTTGTGTTTGTGTTTGTGTTTGTGTT 4440
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 GAGAGAGAGAGAGAGAGAGAGAGAGAGAGAGATCTACCATATAGAAATGATTAGG 4560
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 AATTAAGAGATCTTTTGTATGTATCTACCAATTTCAATAGTAACATTTCTTGTGCCAT 4860
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 AATGTGTATTTTATTTTCCATACAGTAAAGTGTCTCTCTTCTTCTTCTTCTTCTTCT 4980
 AAGTCCCAAGGCAAGAAAGAAACAAAGTAGGACAGAAAGAGAGAGAGAGAGAGAGAG 5040
 AGACAGAGAGAGAGAGTACAGTGGCTTACGAGGAGGAGTGTGATGAGCTCTGATGATCC 5100
 TTCTCTCATGCTGCTGCTGTGAGGATTTGTGCTGATGAGTACGAGGAGTACAGTGA 5160
 GTGTAATACACATGATGAGTGTACCTGATGCTGTGATTTTGTGCTTCTTCTTCTTCT 5220
 TGTGTAAGTTCGCGGCTGTGAGTGTGTTTGTGCTTGTGATGATGAGTATTTTCTGAG 5280
 ACTGTGAATGTGTGAGCGAGCAGCTGAGGACTTCTTATACAGCTATAAGGCACTTCA 5340
 GAGCATTTGTGTTGTGAGCAGCGCGGCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCT 5400
 CTTGCTATGATAAATCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCT 5460
 CCGAGTGTGTTGAGCAGCTGCGGAAACCATGTCTAGAGTCTTCTTCTTCTTCTTCTTCT 5520
 AGCCACTGATGCTGCTGAGTGTGCTGAGCTGAGCTGTGTTGTAACCTCTTCTTCTTCT 5580
 CACCCGCTATTTTCCCACTTCTGCAATCTGCAAGTCTGCAAGTCTGCAAGTCTGCAAG 5640
 CAGTACGCGGAGTTCACGTGATCTTCCGACAGTCCAGTCTGCTGCTTCTTCTTCTTCT 5700
 GAGGATGCTGCTGCAAGACACAGATCATTTTATACAGGATGATAGAGTCTGATGCGGT 5760
 GATCTTCTTCCCAAGCAATCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCT 5820
 AGCTTCCAAAGCTGAACCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCT 5880
 TAAACCAACCAACAAAGCACTACACATCTGCCCAACCAATGTGTTTCTTCTTCTTCT 5940
 TACCAATTAATCCAGCGTGTGTAACCTGTGAGCCCTGATGATGATGATGATGATGAT 6000
 TGCTGCTTGTAGTGTGACTGTGAGTCTCTATGAGTATGATGATGATGATGATGATGAT 6060
 GATTCAGTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCT 6120
 AACACAGTCTTATCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCT 6180
 CTCTATATCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCT 6240
 GATGTGTAATTAATTAATTAATTAATTAATTAATTAATTAATTAATTAATTAATTAAT 6300
 TCCAGTCTTAAAAAAG 6360

B

Sema6D MGFLLLWFCVFLVSLRAVSFPEDEPLNTVDYHSRQYVPRGRPSNESHRLDFQ 60
 SEMA6D *RVF**CAYI*L*M*Q*****N***** 60

Sema6D LMLKIRDTLYIAGRDQVYTVNLNIPQTEVIPSKKLTWRSRQDRENCAMKHKHDECHN 120
 SEMA6D *****M*K*****N***** 120

Sema6D FIKVFPVRNDPMFVFCGTFNPMCRYYRLRTLEYDGEISGLARCPDARQNTVALFAD 180
 SEMA6D *****S***** 180

Sema6D GKLYSATVADFLASDAVIYRSMGDSALRTIKYDSKWIKEPHFLAIEYGNVYVFFREI 240
 SEMA6D ***** 240

Sema6D AVEHNNLKVAVSRVARICNDMGSGQRVLEKHWTSFLKARLNCNVPDGSFFYFDVLSI 300
 SEMA6D ***** 300

Sema6D TDIIQINGIPTVGVFTTQLNIPGSAVCAFSMDIDIEKVFGRFKEQTPDSVMTAVPED 360
 SEMA6D ***** 360

Sema6D KVPKPRPGCKAKHGLAEAYKTSIDFPDITAFIKSHPLMDSAVPPIADEFPWTKTRVRYR 420
 SEMA6D *****E*S***** 420

Sema6D LTAIEVDRSAGPYQNTYVIFVSGEAGVNLKVLANTSFPFLNSDLVLEEIAYNPACSAE 480
 SEMA6D *****H***** 480

Sema6D SEEDRKVVSLQDKDHALVAFSSCVRLPLSRERYGSCSKSCIASRDYPYCGWLSQGV 540
 SEMA6D *****I***** 540

Sema6D CERVTLGMLLLTEDFFAFHNHSPGQYQDTEYGNTHLGDCHGVREVSQESGNQVHNN 600
 SEMA6D *Q*P*****AE*****P***** 600

Sema6D VLITCVFAFVLGAFIAGVAVYCYRDMFVRKNRKIKHDAESAQSCTDSSGSFALKNLGLF 660
 SEMA6D ***** 660

Sema6D SPVKYQQNIDSKPLYSNLLTSRKELPPNTDTKSHAVDHRGQPPLEALPTPESTPLVHQ 720
 SEMA6D *****V***** 720

Sema6D KTLQAMKSHSEKASHGASRKEHPQFFSPPPPSPHSLSHGHPISAIPLNATHDYNTSFS 780
 SEMA6D ***** 780

Sema6D NSNAHAEKKLGMDHPLTKSSKREHRSVDSENRLNLDLKLHNDPNSNPKAILGEIHM 840
 SEMA6D *****N*****D*****M*****P*****Q* 840

Sema6D AHQTLMLDPVGMAEVFPKVPNREASLYSPPTLPNSPTKRVDVPTTGPVPMTSLSRQR 900
 SEMA6D *****M*S*S***** 900

Sema6D GYHNSQSRHSISAVPKNLNSPGLVLSRQPSNHRGQYMPPTGAKVDYIQGTVPVSHLQ 960
 SEMA6D *****M***** 960

Sema6D PSLRSQSYTNSGTLPRTLGKRTPSLKPDPVPPKPSFVQPTTSVRPLNKYTY 1011
 SEMA6D *****P***** 1011

Fig. 1. Identification of murine Sema6D gene. (A) The nucleotide and predicted amino acid sequences of murine Sema6D-1. Putative signal sequence and transmembrane domain are underlined. The in-frame stop codon and poly(A) signal sequences are shown in bold letters. (B) Alignment of murine Sema6D-1 (Sema6D) and human SEMA6D proteins.

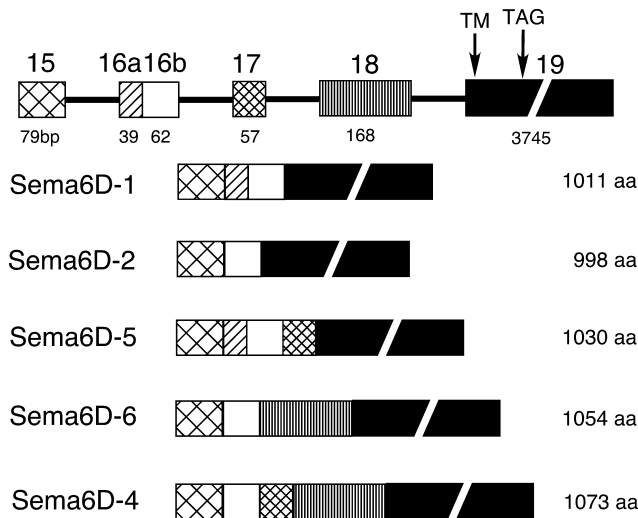


Fig. 2. Schematic diagram of generation of Sema6D isoforms. The boxes and lines represent exons and introns, respectively. The length of introns is arbitrary. The exons 15–19 and the length (bp) of their exons are shown, and the coding exons 2–14 are identical in all isoforms and omitted from the figure. The nomenclature of isoforms was adjusted with human Sema6D [23]. TM represents a putative transmembrane region. The stop codon TAG is also indicated.

Sema6D transcripts were detected in the olfactory bulb, dentate gyrus, and other regions (Figs. 4D and F, shown in blue), suggesting important roles in these regions.

Expression pattern of Sema6D isoforms

To clarify expression profile of Sema6D isoforms in various tissues, RT-PCR was performed by a common primer set designed on exons 15 and 19 (see Materials and methods) (Fig. 5). Most of the Sema6D isoforms were detected in all tissues, but their expression level differed depending on tissues. It is noteworthy that all isoforms were expressed in the brain, while Sema6D-2 and Sema6D-6 were the major transcripts in other tissues. Thus, it is possible that individual isoforms play both redundant and distinct biological functions.

Discussion

In the present study, we identified and cloned murine Sema6D cDNAs. Predictable protein of Sema6D-1 is 1011 aa and showed a 95.3% identity with that of human SEMA6D (KIAA1479). We also identified four Sema6D isoforms. These isoforms were generated by alternative splicing (Fig. 2). In all other class 6 semaphorins, isoforms were reported [19,22,24,25]. Sema6C isoforms are generated by alternative splicing and the isoform expression is controlled in a tissue- and stage-dependent manner, though functional differences between the isoforms have not been reported [22]. We found that all

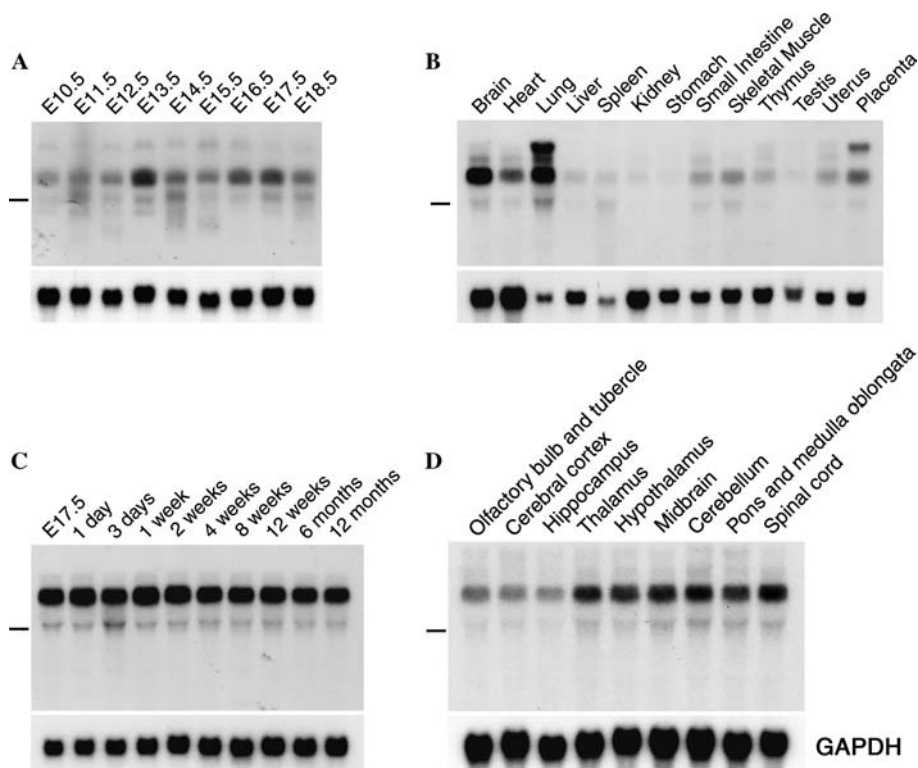


Fig. 3. Analysis of Sema6D expression by Northern blotting. Each lane contained 20 μ g of total RNA. (A) Sema6D expression of the whole embryo during murine embryogenesis. (B) The distribution of Sema6D in adult murine tissues. (C) Sema6D expression in murine brain at various developmental stages. (D) Regional distribution of Sema6D in adult rat brain. The lines show the location of murine 28S ribosomal RNA. Glyceraldehyde-3-phosphate dehydrogenase (GAPDH) expression patterns are also shown as references.

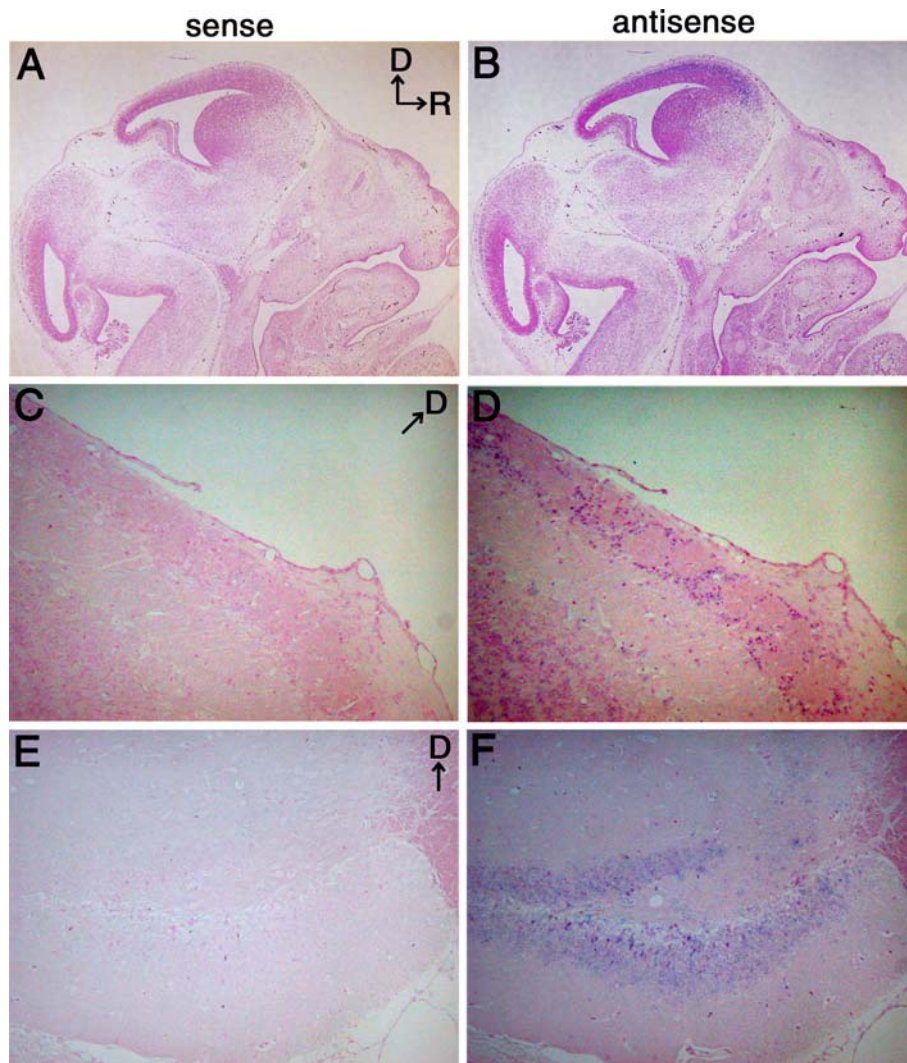


Fig. 4. Sema6D expression pattern of sagittal sections by in situ hybridization with sense riboprobe (A,C,E) and antisense riboprobe (B,D,F). (A,B) are from murine E13.5 and (C–F) are from adult murine brain. (B) At E13.5 Sema6D was expressed highly in the neopallial cortex (shown in blue). (D,F) In adult brain Sema6D transcripts were detected in the glomerular layer of the olfactory bulb and dentate gyrus (shown in blue). D and R represent dorsal and rostral, respectively.

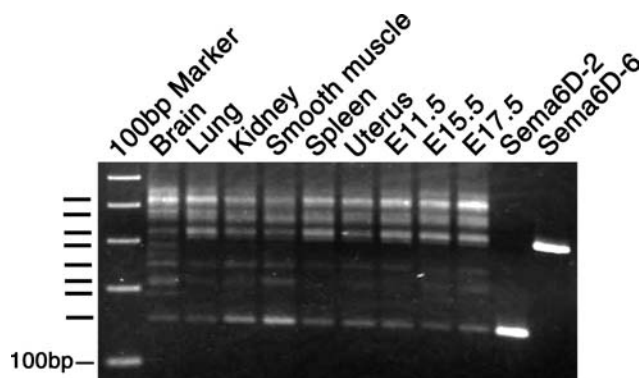


Fig. 5. Expression pattern of Sema6D isoforms. RT-PCR was performed with PCR primers designed on exons 15 and 19. Putative isoform bands are shown by thick bars at the left. The sizes of Sema6D isoforms are as follows: Sema6D-2 (155 bp), Sema6D-1 (194 bp), Sema6D-5 (251 bp), Sema6D-6 (323 bp), and Sema6D-4 (380 bp). Several unidentified bands are also seen.

Sema6D isoforms were identified in adult brain (Fig. 5). Northern blot analysis showed that in brain Sema6D transcript exhibits a single band (Fig. 3), suggesting that either the lengths of Sema6D isoform transcripts are similar to that of Sema6D-1 transcript, or the content of other variants is minute. To clarify the expression profile of individual transcripts, RT-PCR was performed with various tissues (Fig. 5). In the brain all isoforms are expressed, while Sema6D-2 and Sema6D-6 are predominant transcripts in other tissues. These results suggest that the alternative splicing is regulated in a tissue-specific manner, although the biological function of each isoform remains to be revealed.

Sema6D was expressed during embryogenesis (Fig. 3A), suggesting that Sema6D may function in neurogenesis and morphogenesis like Sema3A [6,26,27]. It was expressed predominantly in the brain and its

ubiquitous expression in the brain continued from embryo to adulthood (Figs. 3B, C, and D). Thus, *Sema6D* may function in neurogenesis and plasticity in the brain. It was also expressed in the lung (Fig. 3B) like other semaphorins, but the function of semaphorins in the lung has not been analyzed except for *Sema3A* [28]. *Sema6D* was expressed in the heart and placenta. *Sema3A* and *Sema3C* function during cardiac development [17,26]. Class 3 semaphorins control vascular morphogenesis [29]. In fact, neuropilin-1 is a class 3 semaphorin receptor and also a vascular endothelial growth factor (VEGF) co-receptor [30], functioning in vasculogenesis [31]. Plexin-A2 is a component of a *Sema3A* receptor complex with Neuropilin-1 [14,32] and may function during cardiac development [33]. Thus, semaphorins and their respective receptors play roles in vasculogenesis and angiogenesis besides the axon guidance.

During the preparation of this manuscript, human *SEMA6D* was reported independently [23]. They showed that human *SEMA6D* induced the growth cone collapse of DRG and hippocampal neurons in culture. The information of murine gene would be useful for further functional analysis of this molecule in vivo.

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